

Supporting Information for:

Chemical Gradients within Brain Extracellular Space Measured using Low Flow Push-Pull
Perfusion Sampling *in Vivo*

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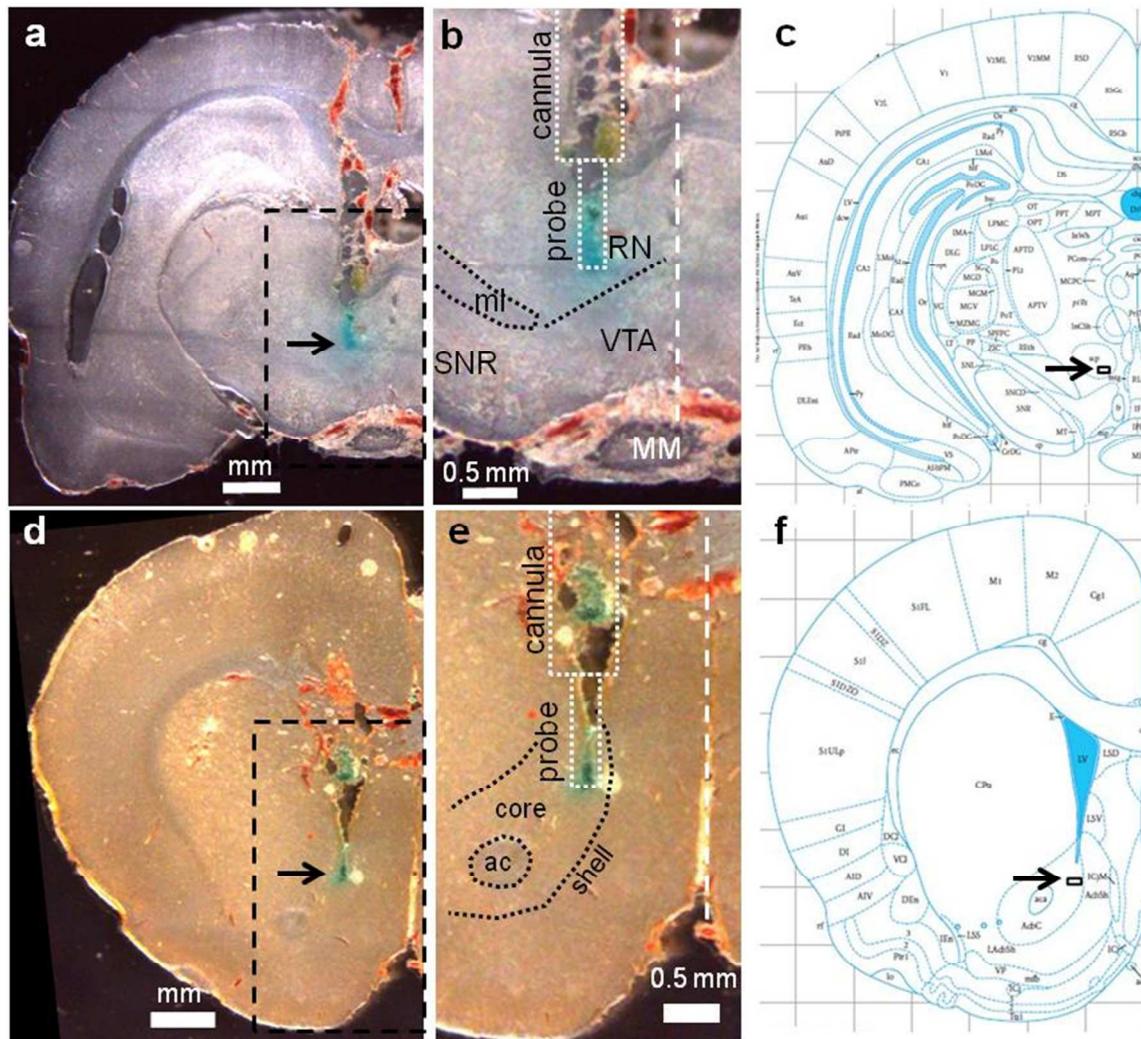


Figure 1. Examples of histology for probe tracking. Coronal brain slices were mounted to microscope slides (**a, d**). Fast Green FCF stained the probe track a blue color, whereas the cannula track was inherently visible by displaced tissue (**a, b, d, e**). Anatomical features including white matter, ventricles, and regional boundaries provided references for identifying placements (**b, e**). Probe tip placements were then mapped on diagrams of coronal slices (**c, f**). Panels **b** and **e** include overlays of the probe and cannula locations (drawn to scale), the medial line (white), and visible features. Arrows indicate sampling sites (**a, c, d, e**). Abbreviations: SNR – substantia nigra reticulata; ml – medial lemniscus; MM – medial mammillary nucleus; ac – anterior commissure; RN – red nucleus; VTA – ventral tegmental area.

Region	Neurotransmitter	“No-Net-Flux” Concentration	Species	Push-Pull Concentration
VTA	DA	5.0 ± 0.5 nM(2)	♀ Indiana “P”	4.8 ± 1.5 nM
	Glu	3.3 ± 0.8 µM, 4.1 ± 0.5 µM(3)	♀ Wistar	8.5 ± 2.7 µM
Accumbens	5-HT	0.7 nM(4)	♂ SD	1.3 ± 0.6 nM (core)
		0.7 ± 0.1 nM(5)	♂ Wistar	0.6 ± 0.2 nM (shell)
		0.6 ± 0.1 nM(6)	♂ Wistar	
	DA	4.7 ± 0.7 nM(7)	♂ Holtzman	7.2 ± 1.2 nM (core)
		5.6 ± 0.4 nM(5)	♂ Wistar	11 ± 4 nM (shell)
		8.3 ± 1.2 nM(6)	♂ Wistar	
	GABA	32.7 ± 4.0 nM(8)	♂ SD	150 ± 110 nM (core) 92 ± 48 nM (shell)
		1.8 ± 0.4 µM, 2.4 ± 0.5 µM(9)	♂ SD	2.4 ± 0.9 µM (core)
	Glu	5.6 ± 1.0 µM(10)	♂ SD	0.93 ± 0.34 µM (shell)
Striatum (anesthetized)	DA	2.5 ± 0.5 nM(11)	♂ SD	1.7 ± 0.2 nM
		6.5 ± 1.1 nM(12)	♂ SD	
	Glu	3.0 ± 0.6 µM(13)	♂ SD	1.1 ± 0.2 µM

Table 1. Comparison of concentrations measured by microdialysis calibrated by “no-net-flux”, and low-flow push-pull perfusion (this work). Male Sprague-Dawley (SD) rats were utilized for push-pull, whereas animals in referenced microdialysis studies varied, as shown above.

References:

1. Paxinos, G., and Watson, C. (2008) *The Rat Brain in Stereotaxic Coordinates*, Academic Press, San Diego, CA.
2. Engleman, E. A., Keen, E. J., Tilford, S. S., Thielen, R. J., and Morzorati, S. L. (2011) Ethanol drinking reduces extracellular dopamine levels in the posterior ventral tegmental area of nondependent alcohol-preferring rats, *Alcohol* 45, 549-557.
3. Ding, Z.-M., Engleman, E. A., Rodd, Z. A., and McBride, W. J. (2012) Ethanol Increases Glutamate Neurotransmission in the Posterior Ventral Tegmental Area of Female Wistar Rats, *Alcohol.: Clin. Exp. Res.* 36, 633-640.
4. Tao, R., Ma, Z., and Auerbach, S. B. (2000) Differential Effect of Local Infusion of Serotonin Reuptake Inhibitors in the Raphe versus Forebrain and the Role of Depolarization-Induced Release in Increased Extracellular Serotonin, *J. Pharmacol. Exp. Ther.* 294, 571-579.
5. Smith, A. D., and Weiss, F. (1999) Ethanol Exposure Differentially Alters Central Monoamine Neurotransmission in Alcohol-Preferring versus -Nonpreferring Rats, *J. Pharmacol. Exp. Ther.* 288, 1223-1228.
6. Katner, S. N., and Weiss, F. (2001) Neurochemical Characteristics Associated With Ethanol Preference in Selected Alcohol-Preferring and -Nonpreferring Rats: A Quantitative Microdialysis Study, *Alcohol.: Clin. Exp. Res.* 25, 198-205.
7. Crippens, D., Camp, D. M., and Robinson, T. E. (1993) Basal extracellular dopamine in the nucleus accumbens during amphetamine withdrawal: a 'no net flux' microdialysis study, *Neurosci. Lett.* 164, 145-148.
8. Xi, Z.-X., Ramamoorthy, S., Shen, H., Lake, R., Samuvel, D. J., and Kalivas, P. W. (2003) GABA Transmission in the Nucleus Accumbens Is Altered after Withdrawal from Repeated Cocaine, *J. Neurosci.* 23, 3498-3505.
9. Melendez, R. I., Hicks, M. P., Cagle, S. S., and Kalivas, P. W. (2005) Ethanol Exposure Decreases Glutamate Uptake in the Nucleus Accumbens, *Alcohol.: Clin. Exp. Res.* 29, 326-333.
10. Baker, D. A., McFarland, K., Lake, R. W., Shen, H., Tang, X.-C., Toda, S., and Kalivas, P. W. (2003) Neuroadaptations in cystine-glutamate exchange underlie cocaine relapse, *Nat Neurosci* 6, 743-749.
11. Chen, N. N. H., Lai, Y.-J., and Pan, W. H. T. (1997) Effects of different perfusion medium on the extracellular basal concentration of dopamine in striatum and medial prefrontal cortex: a zero-net flux microdialysis study, *Neurosci. Lett.* 225, 197-200.
12. Sam, P. M., and Justice, J. B. (1996) Effect of General Microdialysis-Induced Depletion on Extracellular Dopamine, *Anal. Chem.* 68, 724-728.
13. Miele, M., Berners, M., Boutelle, M. G., Kusakabe, H., and Fillenz, M. (1996) The determination of the extracellular concentration of brain glutamate using quantitative microdialysis, *Brain Res.* 707, 131-133.